WB-Validated GOT2 Knockdown Cell Lysate Kit



Catalog #: L62249

Aliases

GOT2; Glutamic-Oxaloacetic Transaminase 2; KYAT4; Kynurenine Aminotransferase IV; MAspAT; KATIV; KAT4; Plasma Membrane-Associated Fatty Acid-Binding Protein; Glutamic-Oxaloacetic Transaminase 2, Mitochondrial; Aspartate Aminotransferase, Mitochondrial; Kynurenine--Oxoglutarate Transaminase 4; Glutamate Oxaloacetate Transaminase 2; Kynurenine Aminotransferase 4; Aspartate Aminotransferase 2; Fatty Acid-Binding Protein; Aspartate Transaminase 2; Transaminase A; EC 2.6.1.1; MitAAT; FABP-1; FABPpm; Glutamic-Oxaloacetic Transaminase 2, Mitochondrial (Aspartate Aminotransferase 2); EC 2.6.1.7; EC 2.6.1; MITAAT; DEE82

Background

Gene Name: GOT2 NCBI Gene Entry: 2806

Storage

Stored at -20°C for 2 years.

Kit Components

1. 100 µg WT cell lysate

2. 100 µg KD cell lysate

Tested Cell Line

HeLa.

Validation Methods

RT-qPCR; Western Blotting (WB)

Shipping

Shipped with gel ice packs. Immediately store the product in a standard freezer at -20°C upon receipt.

Instructions For Use

This knockdown cell lysate should be paired with wild-type HeLa cell lysate for use. For Western blotting, we recommend running wild-type and knockdown lysates on the same gel, and loading each well with equal volume and equal amount of total proteins.

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Manufacturing Process

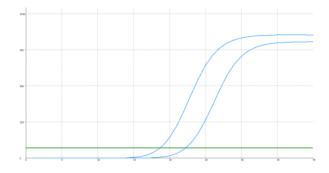
The following protocol was used to generate mRNA knockdown cell lysate:

- 1.Release 0.5 million HeLa cells into a 35 mm tissue culture dish in 2 mL of the growth medium (DMEM containing 10% FBS and 1% pen/strep). Cell density should reach 50-60% confluence the following day.
- 2.24 h after cell release, pre-warm the shRNA lentiviral medium to 37°C.
- 3.Discard 1 mL of the original growth medium of the 35 mm dish.
- 4. Using a serological pipette, gently mix the lentiviral solution 3 times.
- 5.Carefully add 1 mL of the lentiviral solution to the well. Tip: To prevent splashing, add the solution to the dish along the wall.
- 6.Add a polybrene stock solution to the culture medium at a final concentration of 5 $\mu g/mL$. Gently swirl the dish to mix.
- 7.48 h after cell release, without discarding the original medium, add another 1 mL of lentiviral medium directly into the dish.
- 8.Add an additional polybrene stock solution into the dish to obtain a final concentration of 5 μ g/mL. Tip: Now, the medium in the dish should be a total of 3 mL.
- 9.72 h after cell release, cells may reach confluence. Trypsinize the cells off the 35 mm dish and culture those cells in a 60 mm dish.
- 10.Add puromycin to the dish at a final concentration of 4 μ g/mL. Tip: To assess the efficacy of puromycin selection, culture a dish of wild-type HeLa cells as a negative control.
- 11.Allow puromycin selection for 48 h. Almost all wild-type HeLa cells should die, while the dish infected with lentiviruses should have some remaining cells.
- 12.Replace the medium with regular growth medium without puromycin and allow the cells to grow to confluence before harvesting or staining.
- 13.Cells were lysed with IntactProtein™ cell/tissue lysis kit (Cat#415) and stored in -20°C.

Note: This product is for research use only.

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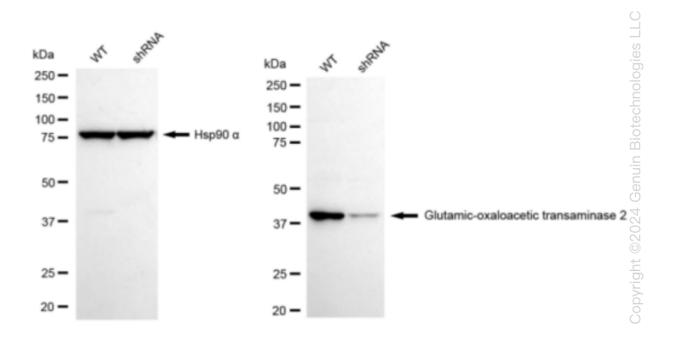
Validation Data



Genotype	Ct Value
Wild-Type	18.42
Knock-Down	21.91
$\Delta Ct (Ct_{KD}-Ct_{WT})$	3.49
% mRNA Reduction	↓ 91%

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RT-qPCR analysis. HeLa cells were infected with GOT2-specific shRNA lentiviral particles, total RNA was extracted from wild-type and knockdown cells, RT-qPCR was performed using gene-specific primers. Δ Ct (CtKD-CtWT) was used to calculate mRNA reduction (%) between wild-type and knockdown cells using the following formula: $(1-1/2\Delta$ Ct) x 100%.



Western blotting analysis. GOT2 protein expression in wild-type (WT) and shRNA knockdown (KD) HeLa cells was detected using Western blotting. Hsp90 α served as a loading control. The blots were incubated with primary antibodies (Cat#62249, 1:5,000) against GOT2 and Hsp90 α, respectively, followed by incubating with HRP-conjugated goat anti-rabbit secondary antibody (Cat#201, 1:20,000). Images were developed using FeQTM ECL Substrate Kit (Cat#226).