

WB-Validated HSPD1 Lentiviral shRNA Knockdown Kit



Catalog #: V61476

Aliases

HSPD1; Heat Shock Protein Family D (Hsp60) Member 1; HSP60; GroEL; 60 KDa Heat Shock Protein, Mitochondrial; Heat Shock 60kDa Protein 1 (Chaperonin); Heat Shock Protein Family D Member 1; Mitochondrial Matrix Protein P1; P60 Lymphocyte Protein; 60 KDa Chaperonin; Chaperonin 60; HuCHA60; HSP-60; CPN60; SPG13; Spastic Paraplegia 13 (Autosomal Dominant); Epididymis Secretory Sperm Binding Protein; Heat Shock 60kD Protein 1 (Chaperonin); Short Heat Shock Protein 60 Hsp60s1; Heat Shock Protein 65; Heat Shock Protein 60; EC 5.6.1.7; HSP65; Hsp60; HLD4

Background

Gene Name: HSPD1

NCBI Gene Entry: [3329](#)

Storage

Store at -80 °C for one year.

Kit Components

1. WB-Validated HSPD1 shRNA lentiviral particles (4 mL)
2. Non-Target shRNA lentiviral particles (4 mL)
3. Verification Tool: KD-Validated Anti-Hsp60 Rabbit mAb #61476 (5 µL)

Tested Cell Line

HeLa

Validation Methods

RT-qPCR; Western Blotting (WB)

Shipping

Shipped with dry ice. Immediately store the product in a standard freezer at -80°C upon receipt.

Instructions For Use

The following protocol uses HeLa cell as an example assuming your cell culture medium is DMEM-based.

1. Release 0.5 million HeLa cells into a 35 mm tissue culture dish in 2 mL of the growth medium (DMEM containing 10% FBS and 1% pen/strep). Cell density should reach 50-60% confluence the following day.

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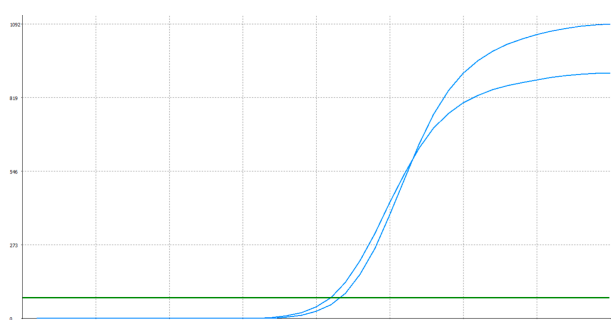
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2. 24 h after cell release, pre-warm the shRNA lentiviral medium to 37°C.
3. Discard 1 mL of the original growth medium of the 35 mm dish.
4. Using a serological pipette, gently mix the lentiviral solution 3 times.
5. Carefully add 1 mL of the lentiviral solution to the well. Tip: To prevent splashing, add the solution to the dish along the wall.
6. Add a polybrene stock solution to the culture medium at a final concentration of 5 µg/mL. Gently swirl the dish to mix.
7. 48 h after cell release, without discarding the original medium, add another 1 mL of lentiviral medium directly into the dish.
8. Add an additional polybrene stock solution into the dish to obtain a final concentration of 5 µg/mL. Tip: Now, the medium in the dish should be a total of 3 mL.
9. 72 h after cell release, cells may reach confluence. Trypsinize the cells off the 35 mm dish and culture those cells in a 60 mm dish.
10. Add puromycin to the dish at a final concentration of 4 µg/mL. Tip: To assess the efficacy of puromycin selection, culture a dish of wild-type HeLa cells as a negative control.
11. Allow puromycin selection for 48 h. Almost all wild-type HeLa cells should die, while the dish infected with lentiviruses should have some remaining cells.
12. Replace the medium with regular growth medium without puromycin and allow the cells to grow to confluence before harvesting or staining.

- Note:**
1. This product is for research use only.
 2. This product is only supplied to end users.
 3. Do not use this product for commercial.

Validation Data



Genotype	Ct Value
Wild-Type	20.63
Knock-Down	21.61
$\Delta C_t (C_{t_{KD}} - C_{t_{WT}})$	0.98
% mRNA Reduction	↓ 49%

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RT-qPCR analysis. HeLa cells were infected with HSPD1-specific shRNA lentiviral particles, total RNA was extracted from wild-type and knockdown cells, RT-qPCR was performed using gene-specific primers. $\Delta C_t (C_{t_{KD}} - C_{t_{WT}})$ was used to calculate mRNA reduction (%) between wild-type and knockdown cells using the following formula: $(1 - 1/2^{\Delta C_t}) \times 100\%$.

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