# WB-Validated MITF Lentiviral shRNA Knockdown Kit



**Catalog #: V62865** 

#### **Aliases**

MITF; Melanocyte Inducing Transcription Factor; BHLHe32; Microphthalmia-Associated Transcription Factor; MI; Melanogenesis Associated Transcription Factor; Class E Basic Helix-Loop-Helix Protein 32; WS2A; WS2; Microphtalmia-Associated Transcription Factor; Homolog Of Mouse Microphthalmia; Waardenburg Syndrome, Type 2A; BHLHE32; COMMAD; MITF-A; CMM8

### **Background**

Gene Name: MITF NCBI Gene Entry: 4286

# **Storage**

Store at -80 °C for one year.

# **Kit Components**

- 1. WB-Validated MITF shRNA lentiviral particles (4 mL)
- 2. Non-Target shRNA lentiviral particles (4 mL)
- 3. Verification Tool: KD-Validated Anti-MITF Mouse mAb #62865 (5 µL)

#### **Tested Cell Line**

HeLa

### **Validation Methods**

RT-qPCR; Western Blotting (WB)

### **Shipping**

Shipped with dry ice. Immediately store the product in a standard freezer at -80°C upon receipt.

### **Instructions For Use**

The following protocol uses HeLa cell as an example assuming your cell culture medium is DMEM-based.

1.Release 0.5 million HeLa cells into a 35 mm tissue culture dish in 2 mL of the growth medium (DMEM containing 10% FBS and 1% pen/strep). Cell density should reach 50-60% confluence the following day.

- 2.24 h after cell release, pre-warm the shRNA lentiviral medium to 37°C.
- 3.Discard 1 mL of the original growth medium of the 35 mm dish.

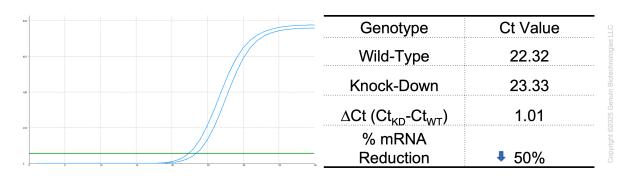
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- 4. Using a serological pipette, gently mix the lentiviral solution 3 times.
- 5.Carefully add 1 mL of the lentiviral solution to the well. Tip: To prevent splashing, add the solution to the dish along the wall.
- 6.Add a polybrene stock solution to the culture medium at a final concentration of 5  $\mu g/mL$ . Gently swirl the dish to mix.
- 7.48 h after cell release, without discarding the original medium, add another 1 mL of lentiviral medium directly into the dish.
- 8.Add an additional polybrene stock solution into the dish to obtain a final concentration of 5  $\mu$ g/mL. Tip: Now, the medium in the dish should be a total of 3 mL.
- 9.72 h after cell release, cells may reach confluence. Trypsinize the cells off the 35 mm dish and culture those cells in a 60 mm dish.
- 10.Add puromycin to the dish at a final concentration of 4  $\mu$ g/mL. Tip: To assess the efficacy of puromycin selection, culture a dish of wild-type HeLa cells as a negative control.
- 11.Allow puromycin selection for 48 h. Almost all wild-type HeLa cells should die, while the dish infected with lentiviruses should have some remaining cells.
- 12.Replace the medium with regular growth medium without puromycin and allow the cells to grow to confluence before harvesting or staining.

**Note:** 1. This product is for research use only.

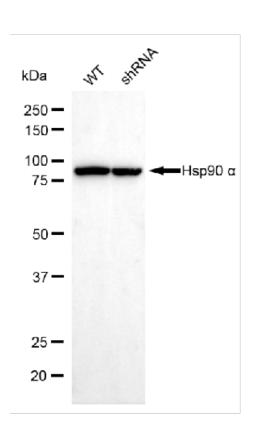
- 2. This product is only supplied to end users.
- 3. Do not use this product for commercial.

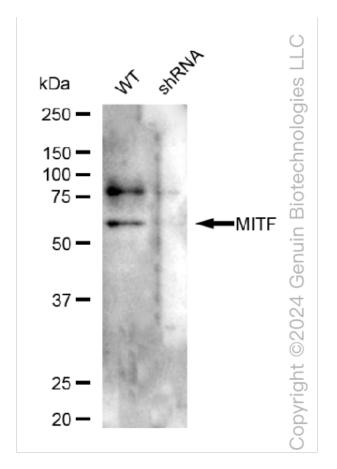
#### Validation Data



RT-qPCR analysis. HeLa cells were infected with MITF-specific shRNA lentiviral particles, total RNA was extracted from wild-type and knockdown cells, RT-qPCR was performed using gene-specific primers.  $\Delta$ Ct (CtKD-CtWT) was used to calculate mRNA reduction (%) between wild-type and knockdown cells using the following formula:  $(1-1/2\Delta$ Ct) x 100%.

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Western blotting analysis. MITF protein expression in wild-type (WT) and shRNA knockdown (KD) HeLa cells was detected using Western blotting. Hsp90  $\alpha$  served as a loading control. The blots were incubated with primary antibodies against MITF and Hsp90  $\alpha$ , respectively, followed by incubating with HRP-conjugated goat anti-mouse secondary antibody. Images were developed using FeQ<sup>TM</sup> ECL Substrate Kit.