# WB-Validated TUBA4A Lentiviral shRNA Knockdown Kit



**Catalog #: V69761** 

#### **Aliases**

TUBA4A; Tubulin Alpha 4a; H2-ALPHA; TUBA1; Tubulin, Alpha 1 (Testis Specific); Tubulin Alpha-4A Chain; Tubulin Alpha-1 Chain; Tubulin H2-Alpha; FLJ30169; Testis-Specific Alpha-Tubulin; Tubulin, Alpha 1; Alpha-Tubulin 1; EC 3.6.5.-; ALS22

## **Background**

Gene Name: TUBA4A NCBI Gene Entry: 7277

## **Storage**

Store at -80 °C for one year.

## **Kit Components**

- 1. WB-Validated TUBA4A shRNA lentiviral particles (4 mL)
- 2. Non-Target shRNA lentiviral particles (4 mL)
- 3. Verification Tool: KD-Validated Anti-TUBA1B/TUBA4A Rabbit mAb #61235 (5 µL)

### **Tested Cell Line**

HT-1080

### Validation Methods

RT-qPCR; Western Blotting (WB)

## **Shipping**

Shipped with dry ice. Immediately store the product in a standard freezer at -80°C upon receipt.

### **Instructions For Use**

The following protocol uses HT-1080 cell as an example assuming your cell culture medium is DMEM-based.

- 1.Release 0.5 million HT-1080 cells into a 35 mm tissue culture dish in 2 mL of the growth medium (DMEM containing 10% FBS and 1% pen/strep). Cell density should reach 50-60% confluence the following day.
- 2.24 h after cell release, pre-warm the shRNA lentiviral medium to 37°C.
- 3.Discard 1 mL of the original growth medium of the 35 mm dish.
- 4. Using a serological pipette, gently mix the lentiviral solution 3 times.
- 5. Carefully add 1 mL of the lentiviral solution to the well. Tip: To prevent splashing, add the

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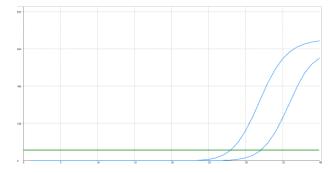
solution to the dish along the wall.

- 6.Add a polybrene stock solution to the culture medium at a final concentration of 5  $\mu$ g/mL. Gently swirl the dish to mix.
- 7.48 h after cell release, without discarding the original medium, add another 1 mL of lentiviral medium directly into the dish.
- 8.Add an additional polybrene stock solution into the dish to obtain a final concentration of 5  $\mu$ g/mL. Tip: Now, the medium in the dish should be a total of 3 mL.
- 9.72 h after cell release, cells may reach confluence. Trypsinize the cells off the 35 mm dish and culture those cells in a 60 mm dish.
- 10.Add puromycin to the dish at a final concentration of 4  $\mu$ g/mL. Tip: To assess the efficacy of puromycin selection, culture a dish of wild-type HT-1080 cells as a negative control.
- 11.Allow puromycin selection for 48 h. Almost all wild-type HT-1080 cells should die, while the dish infected with lentiviruses should have some remaining cells.
- 12.Replace the medium with regular growth medium without puromycin and allow the cells to grow to confluence before harvesting or staining.

**Note:** 1. This product is for research use only.

- 2. This product is only supplied to end users.
- 3. Do not use this product for commercial.

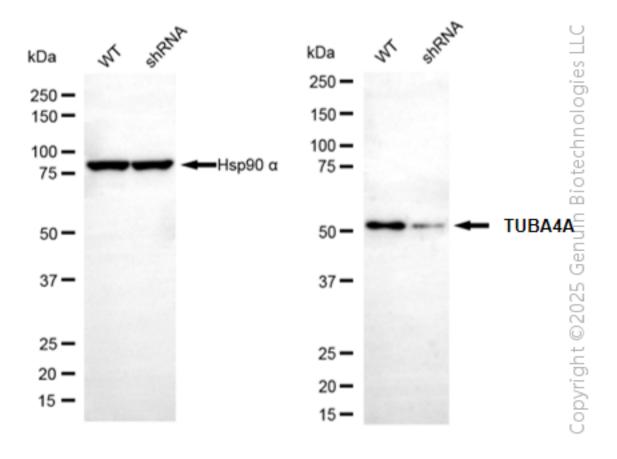
#### Validation Data



Genotype	Ct Value
Wild-Type	27.61
Knock-Down	31.42 iji
ΔCt (CtKD-CtWT)	3.81
% mRNA	Copyright (
Reduction	93% <sup>§</sup>

RT-qPCR analysis. HT-1080 cells were infected with TUBA4A-specific shRNA lentiviral particles, total RNA was extracted from wild-type and knockdown cells, RT-qPCR was performed using gene-specific primers.  $\Delta$ Ct (CtKD-CtWT) was used to calculate mRNA reduction (%) between wild-type and knockdown cells using the following formula:  $(1-1/2\Delta$ Ct) x 100%.

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Western blotting analysis. TUBA4A protein expression in wild-type (WT) and shRNA knockdown (KD) HT-1080 cells was detected using Western blotting. Hsp90  $\alpha$  served as a loading control. The blots were incubated with primary antibodies against TUBA4A and Hsp90  $\alpha$ , respectively, followed by incubating with HRP-conjugated goat anti-rabbit secondary antibody. Images were developed using FeQ<sup>TM</sup> ECL Substrate Kit.